

Damage and Reproduction Potentials of *Heterodera avenae* on Wheat under Outdoor Conditions

A. S. AL-HAZMI, F. A. AL-YAHYA, AND A. T. ABDUL-RAZIG¹

Abstract: Two pot experiments, in consecutive years, were conducted under outdoor conditions during the wheat growing season to examine the relationship between increasing initial population densities (Pi: 0–3,000 cysts/pot) of *Heterodera avenae* and corresponding responses of wheat cv. Yecora Rojo. Results of both experiments were very similar. The nematode suppressed plant height, root and biomass dry weights, and grain yield at all Pi's studied. The suppression of these parameters, as well as the final nematode population densities (Pf), increased with increasing Pi levels. The reproduction factor (Pf/Pi) decreased as Pi increased but was always greater than 1.0. When data from both experiments were combined for regression analyses, inverse relationships were found between $\log_{10}(Pi + 1)$ and both plant growth and yield. These negative relationships were highly significant and adequately described by linear models. Final population (Pf) increased linearly with Pi. The wheat cultivar cv. Yecora Rojo was found to be highly vulnerable to damage and a good host for *H. avenae*.

Key words: cereal cyst nematode, growth, *Heterodera avenae*, nematode, pathogenicity, reproduction, *Triticum aestivum*, wheat, yield.

Wheat (*Triticum aestivum* L.) is the most important field crop in Saudi Arabia. The crop is cultivated, under irrigation, throughout most of the country but most intensively in the Central Province. In 1982, the area under wheat cultivation was 735,000 ha and yield was only 187,000 MT. Since then production has increased to a peak in 1992, when the cultivated area was 924,407 ha producing 4.123 million MT (Anonymous, 1996).

The cereal cyst nematode (CCN), *Heterodera avenae* Wollenweber, was first reported on wheat and barley in the Riyadh region in 1992 (Al-Hazmi, 1992). The nematode pathotype infesting the region is very similar to Ha21 (Cook and Al-Hazmi, 1997). *H. avenae* has been spreading and has become a major limiting factor in wheat production in most wheat-producing regions in the country (Al-Hazmi et al., 1994; Al-Yahya et al., 1998; El-Meleigi and Al-Rokaibah, 1996). Most of the wheat cultivars grown in the country are susceptible to CCN (Al-Hazmi et al., 1994). Severe symptoms of stunting and yellowing often occur, and the nematode density in some heavily infested fields may reach 67 second-stage juveniles

(J2)/g soil (Al-Hazmi et al., 1994), and yield loss may be as high as 93.5% (Al-Yahya et al., 1996).

Although previous reports or observations have provided evidence of CCN damage to wheat in Saudi fields, experimental studies are still needed to establish the damage function and reproductive potential of *H. avenae* on wheat. The objective of this study was to examine the relationship between increasing population densities (Pi) of *H. avenae* and the corresponding responses of wheat cv. Yecora Rojo in outdoor pot experiments.

MATERIALS AND METHODS

Clean 12-cm-diam. plastic pots were filled with a steam-sterilized soil mixture of equal parts of sand and loamy sand soil. The soil mixture of each pot (ca. 1,500 cm³ soil) was mixed thoroughly in a polyethylene bag with the designated inoculum of *H. avenae* and returned to its pot. The inoculum (brown cysts) was obtained from soil collected at the end of the previous season from a heavily infested wheat field in Al-Kharj region near Riyadh and stored at 5 °C. Cysts were extracted from soil with the modified Cobb decanting and sieving method (Barker, 1985). The average number of infective second-stage juveniles (J2) per cyst, determined from a sample of 50 cysts, was 53.5 J2/cyst. Cysts were used as inoculum,

and six initial inoculum levels (Pi) 375, 750, 1,500, and 3,000 cysts/pot were used. These densities correspond to 10,000, 20,000, 40,000, 80,000, and 160,000 J2/pot. Treatments were replicated five times. The control treatment received the same volume of cyst-free decanting solution. Four uniform seedlings of wheat cv. Yecora Rojo, a susceptible cultivar (Al-Hazmi et al., 1994), were transplanted into each pot. Plants were watered after transplanting and arranged in a randomized complete block design on a cardboard outside the greenhouse during the 1996/1997 wheat growing season. As needed, plants were watered and fertilized with water-soluble N-P-K fertilizer. The experiment was repeated the following season (1997/1998).

At harvest in both years (120 days after transplanting), plant heights were measured, and grain weight per spikelet per pot were determined. Roots were gently from soil and weighed. Cysts were dislodged by first washing roots to free them from soil, and then watered with a strong jet of water onto a 850-µm sieve over a 150-µm-pore sieve (Barker, 1986). Soil from each pot was thoroughly mixed, and a 250-cm³ aliquot was used to extract cysts and J2 with a modified decantation procedure (Barker, 1985) in a dense sucrose solution (Dunn, 1966). The numbers of J2 found in the soil and roots were negligible, final population density was calculated from cysts collected from roots and soil. The reproduction factor (Pf/Pi) for each replicate was calculated.

TABLE 1. Growth responses of wheat cv. Yecora Rojo to different inoculum levels of *H. avenae* in two outdoor pot experiments.

Pi (cysts/1,500 cm ³ soil)	Plant height (cm)	
	1996	1997
0	45.2 a	52.1
187	40.7 b	42.4
375	33.7 c	38.4
750	29.2 d	34.6
1,500	27.4 d	28.6
3,000	24.8 e	26.0

Values are means of five replicates. Means in a column are significantly different according to Fisher's Protected LSD.

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¹ Professor, Assistant Professor, and Research Assistant, Department of Plant Protection, King Saud University, Box 2460, Riyadh 11451, Saudi Arabia.

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Effects of *Heterodera avenae* on Wheat under Outdoor Conditions

A. T. ABDUL-RAZIG¹

Conducted under outdoor conditions during 1996/1997 and 1997/1998, the effects of increasing initial population densities of *H. avenae* on the growth responses of wheat cv. Yecora Rojo. Parameters measured included plant height, root and biomass dry weights, and total biomass. The reproduction factor (Rf) was calculated from the final population density (Pf) and the initial population density (Pi). The wheat cultivar cv. Yecora Rojo was found to be highly susceptible to *H. avenae*. The nematode, pathogenicity, reproduction,

and yield (Al-Hazmi et al., 1994), and yield loss were as high as 93.5% (Al-Yahya et al., 1994).

Although previous reports or observations have provided evidence of CCN damage to wheat in Saudi fields, experimental studies were needed to establish the damage caused by *H. avenae* and its reproductive potential of *H. avenae* on wheat. The objective of this study was to examine the relationship between initial population densities (Pi) of *H. avenae* and the corresponding responses of wheat cv. Yecora Rojo in outdoor pot experi-

MATERIALS AND METHODS

12-cm-diam. plastic pots were filled with steam-sterilized soil mixture of equal parts of sand and loamy sand soil. The soil in each pot (ca. 1,500 cm³ soil) was sterilized thoroughly in a polyethylene bag and designated inoculum of *H. avenae* was added to its pot. The inoculum (J2 cysts) was obtained from soil collected from the end of the previous season from a heavily infested wheat field in Al-Riyadh near Riyadh and stored at 5 °C. The nematodes were extracted from soil with the modified decanting and sieving method (Hooper, 1985). The average number of second-stage juveniles (J2) per cyst, determined from a sample of 50 cysts, was used as inoculum.

and six initial inoculum levels (Pi) of 0, 187, 375, 750, 1,500, and 3,000 cysts/pot were used. These densities correspond to 0, 10,000, 20,000, 40,000, 80,000, and 160,000 J2/pot. Treatments were replicated five times. The control treatment received the same volume of cyst-free decanting water. Four uniform seedlings of wheat cv. Yecora Rojo, a susceptible cultivar (Al-Hazmi et al., 1994), were transplanted into each pot. Pots were watered after transplanting and arranged in a randomized complete block design on a cardboard outside the greenhouse during the 1996/1997 wheat growing season. As needed, plants were watered and fertilized with water-soluble N-P-K (20-20-20) fertilizer. The experiment was repeated in the following season (1997/1998).

At harvest in both years (120 or 130 days after transplanting), plant heights were measured, and grain weight per spike and per pot were determined. Roots were separated gently from soil and weighed. Cysts on roots were dislodged by first washing roots gently to free them from soil, and then vigorously with a strong jet of water onto a 850-µm-pore sieve over a 150-µm-pore sieve (Hooper, 1986). Soil from each pot was thoroughly mixed, and a 250-cm³ aliquot was taken to extract cysts and J2 with a modified centrifugation procedure (Barker, 1985) and a dense sucrose solution (Dunn, 1969). Since the numbers of J2 found in the soil at harvest were negligible, final population (Pf) was calculated from cysts collected from roots and soil. The reproduction factor (Rf = Pf/Pi) for each replicate was calculated. The

shoot and root systems were oven-dried at 60 °C for a week, and dry weights of shoots, roots, and total biomass were determined.

Data from each experiment were subjected to analysis of variance (ANOVA), and treatment means were separated with Fisher's Protected LSD at $P \leq 0.05$ (SAS Institute, Cary, NC). Data from both experiments were combined and regression analyses were performed on selected plant responses and nematode reproduction versus Pi, $\log_{10} (Pi + 1)$, or $\log_{10} Pi$.

RESULTS

Results from both experiments were very similar. Plant heights and dry weights of both roots and biomass were suppressed ($P \leq 0.05$) even at the lowest Pi (187 cysts/pot) (Table 1). Suppression increased with Pi and reached up to 50% in plant height, 81% in roots, and 80% in biomass dry weights at the highest Pi level (3,000 cysts/pot) (Table 1). Similarly, yield (grain weight per pot or grain weight per spike) decreased ($P \leq 0.05$) in response to increasing inoculum level (Table 2). The yield reduction (18-35%) was evident at the lowest Pi, but the greatest reduction (80%) occurred at the highest Pi (Table 2).

Final nematode population (Pf) increased ($P \leq 0.05$) with increasing Pi in both years, but the nematode reproduced more in the second than in the first year (Table 3). Reproduction factors (Rf) decreased ($P \leq 0.05$) in both years, except at Pi = 375 in the second year, as Pi increased

TABLE 1. Growth responses of wheat cv. Yecora Rojo to different initial population densities (Pi) of *Heterodera avenae* in two outdoor pot experiments.

Pi (cysts/1,500 cm ³ soil)	Plant height (cm)		Root dry weight (g)		Biomass dry weight (g)	
	1996	1997	1996	1997	1996	1997
0	45.2 a	52.1 a	1.25 a	1.16 a	7.90 a	7.73 a
187	40.7 b	42.4 b	0.55 b	0.58 b	7.02 b	5.80 b
375	33.7 c	38.4 c	0.41 c	0.46 c	4.93 c	4.49 c
750	29.2 d	34.6 d	0.26 d	0.34 d	3.78 d	3.94 d
1,500	27.4 d	28.6 e	0.33 cd	0.27 de	2.85 e	3.44 d
3,000	24.8 e	26.0 f	0.29 cd	0.22 e	2.06 f	2.72 e

Values are means of five replicates. Means in a column followed by the same letter are not significantly different ($P \leq 0.05$) according to Fisher's Protected LSD.

TABLE 2. Effect of different initial population densities (Pi) of *Heterodera avenae* on yield of wheat cv. Yecora Rojo in two outdoor pot experiments.

Pi (cysts/1,500 cm ³ soil)	Grain weight/pot (g)		Grain weight/spike (g)	
	1996	1997	1996	1997
0	3.70 a	3.20 a	0.93 a	0.80 a
187	2.41 b	2.64 b	0.60 b	0.66 b
375	1.85 c	1.95 c	0.46 c	0.49 c
750	1.66 c	1.52 d	0.41 c	0.38 d
1,500	1.14 d	0.98 e	0.29 d	0.24 e
3,000	0.74 e	0.73 e	0.19 e	0.18 e

Values are means of five replicates. Means in a column followed by the same letter are not significantly different ($P \leq 0.05$) according to Fisher's Protected LSD.

(Table 3), but Rf was always greater than one.

When data from both growing seasons were combined in the analysis, inverse relationships were found between inoculum level and wheat growth and yield. These negative relationships were highly significant ($P \leq 0.0001$), and adequately described by linear models (Table 4). Plant height (cm) and dry weight (g) of plant biomass were negatively correlated with $\log_{10} (Pi + 1)$ with $R^2 = 0.78$ and 0.80 , respectively. Similarly, yield (grams per pot) was also negatively correlated with $\log_{10} (Pi + 1)$ with $R^2 = 0.83$. Final cyst populations increased linearly with Pi ($R^2 = 0.82$), but the reproduction factor (Rf) was negatively correlated with $\log_{10} (Pi)$ with $R^2 = 0.30$ (Table 4).

DISCUSSION

Heterodera avenae caused significant damage and reproduced well on wheat cv. Yecora Rojo.Suppressions of growth and grain yield were dependent on Pi levels. Even at the lowest Pi of 187 cysts/pot (7 J2/cm³ soil), *H. avenae* caused significant growth and yield suppression. At the highest Pi of 3,000 cysts/pot (107 J2/cm³ soil), grain yield was reduced by up to 80%. Consequently, cultivation of wheat would not be profitable at this Pi level, and greater damage and crop losses might be expected under stressful field conditions. For example, in small field plots, naturally infested with *H. avenae*, we have found that the yield loss of

wheat cv. Yecora Rojo was 93.5% (Al-Yahya et al., 1996). According to Swarup and Sosa-Moss (1990), yield losses of wheat and barley may be 45–48% in light soils even at a density of 6 eggs/g soil. In the Central Province of Saudi Arabia many wheat fields are heavily infested with *H. avenae*, and damage in some fields is so great that some growers plow the soil and grow alfalfa or vegetable crops instead (Al-Hazmi et al., 1994). Although yield depends on many interacting factors, *H. avenae* disturbs several physiological aspects of the infected wheat plants, such as photosynthesis, mineral uptake, transpiration, temperature of the plant canopy, and water content of leaves and roots (Al-Yahya et al., 1998). Our results on suppression of growth and yield of wheat caused by *H. avenae* support different reports from other countries (Brown, 1982; Dhawan and Nagesh, 1987; Greco et al., 1993; Rivoal and Sarr, 1987; Sabova et al., 1986; Williams and Beane, 1982).

In our study, the damage threshold level was 7 J2/g soil (187 cysts/pot). This level is similar to the threshold levels on wheat and barley reported by Swarup and Sosa-Moss (1990) from India (6 eggs/g soil), by Shahina and Maqbool (1990) from Pakistan (100–150 cysts/plant), and by Sabova et al. (1981) from Slovakia (180 cysts/pot). In the temperate semi-arid regions of Australia, which have a somewhat similar climate to that of Saudi Arabia, *H. avenae* reduced yield of wheat and barley by 20% at a Pi of 2 eggs and J2/g soil, and by 40% at a Pi of 16 eggs and J2/g soil (Meagher and Brown, 1974).

TABLE 3. Reproduction of *Heterodera avenae* on wheat cv. Yecora Rojo at different initial population densities (Pi) in two outdoor pot experiments.

Pi (cysts/1,500 cm ³ soil)	Pf (cysts/pot)		Reproduction factor (Rf)	
	1996	1997	1996	1997
187	622 d	744 e	3.32 a	3.97 b
375	733 d	1,884 d	1.95 b	5.02 a
750	1,434 c	2,696 c	1.91 b	3.60 c
1,500	2,464 b	4,273 b	1.64 bc	2.85 d
3,000	4,786 a	8,268 a	1.60 c	2.76 d

Values are means of five replicates. Means in a column followed by the same letter are not significantly different ($P \leq 0.05$) according to Fisher's Protected LSD.

Reproduction factor (Rf) = Pf/Pi.

TABLE 4. Summary of relationships between $\log_{10} (Pi + 1)$ and nematode reproduction (y) on wheat

X	y
$\log_{10} (Pi + 1)$	Plant height (cm)
$\log_{10} (Pi + 1)$	Dry biomass (g/pot)
$\log_{10} (Pi + 1)$	Grain yield (g/pot)
Pi	Pf
$\log_{10} Pi$	Rf

* Data were combined from two outdoor pot experiments (1996 and 1997). Rf = Pf/Pi.

Although several investigators have reported varying damage threshold levels of *H. avenae* on wheat and barley, generally 1 to 10 J2/g soil and J2/g soil have been reported in different countries (Dhawan and Swarup, 1987; Gill and Swarup, 1971; Greco and Brandonisio, 1987; Meagher and Brown, 1974; Stone, 1968; Swarup and Sosa-Moss, 1990). Many biotic and abiotic factors in different regions may affect the establishment of the damage thresholds of *H. avenae* on wheat or other cereals. These factors include: soil type, rainfall, cereal plant cultivar, nematode pathotype, nutrient availability, or location of test, presence of other nematodes or pathogens, and nematicide treatments (Rivoal and Cook, 1993; Simons and Williams and Beane, 1982).

Heterodera avenae reproduced well on wheat cv. Yecora Rojo. Final number of nematode (Pf) increased with Pi ($P = 0.0001$), and the reproduction factor (Rf) was always greater than 1.0. The cultivar was a good host for *H. avenae* but relatively less suitable than some other cultivars and lines (Al-Hazmi et al., 1994), previously reported (Dhawan and Swarup, 1987; O'Brien and Fisher, 1978; Rivoal and Sarr, 1987), Rf was inversely related to Pi ($r = -0.55$, $P = 0.001$). The decrease in Rf is most likely due to the greater damage to roots with increasing inoculum level.

LITERATURE CITED

Al-Hazmi, A. S. 1992. Status of plant nematodes in Saudi Arabia. P. 70 in M. A. Maqbool and others. Expert consultation on plant nematology and their control in the near east region.

Yecora Rojo was 93.5% (Al-Yahya et al. 1994). According to Swarup and Sosa-Moss (1987), yield losses of wheat and barley were 48% in light soils even at a density of 2 eggs/g soil. In the Central Province of Saudi Arabia many wheat fields are heavily infested with *H. avenae*, and damage to wheat is so great that some growers do not plant wheat and grow alfalfa or vegetable crops instead (Al-Hazmi et al., 1994). The extent of damage depends on many interacting factors. *H. avenae* disturbs several physiological processes of the infected wheat plants, such as photosynthesis, mineral uptake, transpiration, temperature of the plant canopy, and growth of leaves and roots (Al-Yahya et al. 1994). Our results on suppression of yield of wheat caused by *H. avenae* are in agreement with other reports from other countries (Brown, 1982; Dhawan and Nagesh, 1987; Greco et al., 1993; Rivoal and Sarr, 1987; Sabova et al., 1986; Williams and Beane, 1982).

In this study, the damage threshold level was 187 cysts/pot. This level is higher than the threshold levels on wheat and barley reported by Swarup and Sosa-Moss in India (6 eggs/g soil), by Sha-Maqbool (1990) from Pakistan (20 cysts/plant), and by Sabova et al. in Slovakia (180 cysts/pot). In the semi-arid regions of Australia, where there is a somewhat similar climate to Saudi Arabia, *H. avenae* reduced yield of wheat and barley by 20% at a P_i of 2 eggs/g soil, and by 40% at a P_i of 16 eggs/g soil (Meagher and Brown, 1974).

Reproduction of *Heterodera avenae* on wheat cv. Yecora Rojo at different initial population densities in two outdoor pot experiments.

Pf (cysts/pot)		Reproduction factor (Rf)	
1996	1997	1996	1997
622 d	744 e	3.32 a	3.97 b
733 d	1,884 d	1.95 b	5.02 a
1,434 c	2,696 c	1.91 b	3.60 c
2,464 b	4,273 b	1.64 bc	2.85 d
4,786 a	8,268 a	1.60 c	2.76 d

Means of five replicates. Means in a column followed by the same letter are not significantly different ($P \leq 0.05$) according to Fisher's Protected LSD.

Reproduction factor (Rf) = Pf/ P_i .

TABLE 4. Summary of relationships between initial inoculum density (P_i) of *Heterodera avenae* (X) and damage and nematode reproduction (y) on wheat cv. Yecora Rojo, 1996–1998.^a

X	y	Linear regression			
		a	b	R ²	P
$\log_{10} (P_i + 1)$	Plant height (cm)	50.96	-6.54	0.78	0.0001
$\log_{10} (P_i + 1)$	Dry biomass (g/pot)	8.35	-1.5	0.80	0.0001
$\log_{10} (P_i + 1)$	Grain yield (g/pot)	3.68	-0.75	0.83	0.0001
P_i	Pf	438.96	2.02	0.82	0.001
$\log_{10} P_i$	Rf	6.85	-1.39	0.30	0.001

^a Data were combined from two outdoor pot experiments ($n = 10$). P_i and Pf are based on number of cysts per pot (1,500 cm^3 soil). Rf = Pf/ P_i .

Although several investigators reported varying damage threshold levels of *H. avenae* on wheat and barley, generally 1 to 40 eggs and 20–100 g soil have been reported from different countries (Dhawan and Nagesh, 1987; Gill and Swarup, 1971; Greco and Brandonisio, 1987; Meagher and Brown, 1974; Stone, 1968; Swarup and Sosa-Moss, 1990). Many biotic and abiotic factors in different regions may affect the estimates of the damage thresholds of *H. avenae* on wheat or other cereals. These factors include: soil type, rainfall, cereal plant or cultivar, nematode pathotype, nutrients, type or location of test, presence of other nematodes or pathogens, and nematicides used in trials (Rivoal and Cook, 1993; Simon, 1980; Williams and Beane, 1982).

Heterodera avenae reproduced readily on wheat cv. Yecora Rojo. Final numbers of nematode (Pf) increased with P_i ($r = 0.90$, $P = 0.0001$), and the reproductive factor (Rf) was always greater than 1.0 (1.6–5.0). The cultivar was a good host for *H. avenae* but relatively less suitable than some barley cultivars and lines (Al-Hazmi et al., 1994). As previously reported (Dhawan and Nagesh, 1987; O'Brien and Fisher, 1978; Rivoal and Sarr, 1987), Rf was inversely related to $\log_{10} P_i$ ($r = -0.55$, $P = 0.001$). The decrease of Rf is most likely due to the greater damage of roots with increasing inoculum levels.

LITERATURE CITED

Al-Hazmi, A. S. 1992. Status of plant nematology in Saudi Arabia. P. 70 in M. A. Maqbool and M. J. Zaki, eds. Expert consultation on plant nematology problems and their control in the near east region. Second In-

ternational Meeting on Plant Nematology, 22–26 November 1992, Karachi, Pakistan (Abstr.).

Al-Hazmi, A. S., A. A. M. Ibrahim, and A. T. Abdul-Razig. 1994. Occurrence, morphology and reproduction of *Heterodera avenae* on wheat and barley in Saudi Arabia. Pakistan Journal of Nematology 12:117–129.

Al-Yahya, F. A., A. A. Alderfasi, A. S. Al-Hazmi, A. A. M. Ibrahim, and A. T. Abdul-Razig. 1998. Effect of the cereal cyst nematode on growth and physiological aspects of wheat under field conditions. Pakistan Journal of Nematology 16:55–62.

Al-Yahya, F. A., A. S. Al-Hazmi, A. A. M. Ibrahim, and A. A. Alderfasi. 1996. Effect of cereal cyst nematode on wheat yield under field conditions. Abstracts of Seventeenth Annual Meeting of The Saudi Biological Society. 28–30 May 1996. Buraidah, Saudi Arabia.

Anonymous. 1996. Statistical year book. Riyadh, Saudi Arabia: Ministry of Agriculture and Water.

Barker, K. R. 1985. Nematode extraction and bioassays. Pp. 19–35 in K. R. Barker, C. C. Carter, and J. N. Sasser, eds. An advanced treatise on *Meloidogyne*. Vol. II. Methodology. Raleigh, NC: North Carolina State University Graphics.

Brown, R. H. 1982. Cultural practices and their effects on *Heterodera avenae* and grain yield of wheat in Victoria, Australia. EPPO Bulletin 12:477–484.

Cook, R., and A. S. Al-Hazmi. 1997. Characterization of a pathotype of cereal cyst nematode, *Heterodera avenae*, from central Saudi Arabia. Journal of Nematology 20:574 (Abstr.).

Dhawan, S. C., and M. Nagesh. 1987. On the relationship between population densities of *Heterodera avenae*, growth of wheat and nematode multiplication. Indian Journal of Nematology 17:231–236.

Dunn, R. A. 1969. Extraction of cysts of *Heterodera* species from soil by centrifugation in high-density solution. Journal of Nematology 1:7 (Abstr.).

El-Meliegi, M. M., and A. A. Al-Rokaibah. 1996. Survey of wheat diseases in central Saudi Arabia. Bulletin of the Faculty of Agriculture, University of Cairo 47:499–512.

Gill, J. S., and G. Swarup. 1971. On the host range of cereal cyst nematode, *Heterodera avenae*, the causal organism of "molya" disease of wheat and barley in Rajasthan. Indian Journal of Nematology 1:63–67.

Greco, N., and A. Brandonisio. 1987. Investigation on *Heterodera avenae* in Italy. Nematologia Mediterranea 15:225–234.

Greco, N., T. D'Addabbo, A. Brandonisio, and F. Elia. 1993. Damage to Italian crops caused by cyst-forming nematodes. Supplement to the Journal of Nematology 25:836-842.

Hooper, D. J. 1986. Extraction of nematodes from plant material. Pp. 51-58 in J. F. Southey, ed. Laboratory methods for work with plant and soil nematodes. London: Her Majesty's Stationery Office.

Meagher, J. W., and R. H. Brown. 1974. Microplot experiment on the effect of plant hosts on populations of the cereal cyst nematode (*Heterodera avenae*) and on the subsequent yield of wheat. *Nematologica* 20:337-346.

O'Brien, P. C., and J. M. Fisher. 1978. Factors influencing the number of larvae of *Heterodera avenae* within susceptible wheat and barley seedlings. *Nematologica* 24:295-304.

Rivoal, R., and R. Cook. 1993. Nematode pests of cereals. Pp. 259-303 in K. Evans, D. L. Trudgill, and J. M. Webster, eds. Plant-parasitic nematodes in temperate agriculture. Wallingford, UK: CAB International.

Rivoal, R., and E. Sarr. 1987. Field experiments on *Heterodera avenae* in France and implications for winter wheat performance. *Nematologica* 33:460-479.

Sabova, M., B. Valocka, and M. Liskova. 1981. Vplyv *Heterodera avenae* na niektore odrody obilnin v experimentálnych podmienkach. *Ochrana Rostlin* 17:191-197.

Sabova, M., B. Valocka, and M. Liskova. 1986. Škodlivost' *Heterodera avenae* na obilninách. *Ochrana Rostlin* 22:301-308.

Shahina, F., and M. A. Maqbool. 1990. Host range studies of *Heterodera zaeae* and *H. avenae*. *Pakistan Journal of Nematology* 8:79-85.

Simon, A. 1980. A plant assay of soil to assess potential damage to wheat by *Heterodera avenae*. *Plant Disease* 64:917-919.

Stone, L. E. W. 1968. Cereal cyst nematode in spring barley damage assessment 1963-64. *Plant Pathology* 17:145-150.

Swarup, G., and C. Sosa-Moss. 1990. Nematode parasites of cereals. Pp. 109-136 in M. Luc, R. A. Sikora, and J. Bridge, eds. Plant parasitic nematodes in subtropical and tropical agriculture. Wallingford, UK: CAB International.

Williams, T. D., and J. Beane. 1982. Variations in cereal yield losses associated with *Heterodera avenae* in England and Wales. *EPPO Bulletin* 12:485-490.

Reniform Nema Sc

R. T. ROBBINS, I. R.

Abstract: Two hundred eighty-two soybean cultivars from Mississippi were tested in greenhouse experiments with resistance to the reniform nematode. Resistant cultivars Forrest and Hartwig were used as controls. Numbers of reniform nematodes of the numbers reproducing on each cultivar were reported. Cultivars with reproduction ratios less than 1.0 whereas those with greater reproduction ratios were in the relative maturity group (RMG) ≤ 4.4 were found to be resistant. Of the 43 cultivars of RMG 5.5-5.9 were resistant. These data will be useful in help control the reniform nematode.

Key words: *Glycine max*, nematode, *Rotylenchulus reniformis*, soybean, susceptible

The reniform nematode, *Rotylenchulus reniformis*, was first described in 1907 on waii (Linford and Oliveira, 1940). It has long been considered primarily a cotton nematode pest (Heald and Thammachar, 1982). *Rotylenchulus reniformis* has a wide host range with at least 314 hosts out of 360 evaluated, including soybean and cotton (Robinson et al., 1997). Since 1907, the reniform nematode has spread throughout much of the eastern half of the U.S. Cotton Belt (Heald and Thammachar, 1982; 1990). Although the greatest incidence appears to be along the Gulf Coast, in the southern part of the Belt, the nematode apparently has the ability to survive in temperate climates, and *R. reniformis* has been associated with cotton as far north as the Lubbock, Texas area (Heald and Thammachar, 1982) and the Missouri bootheel (Robinson et al., 1992). During the past 10 years, the incidence of *R. reniformis* has increased

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² Professor and Research Assistants, Department of Plant Pathology, Nematology Laboratory, University of Arkansas, Fayetteville, AR, 72701.

³ Director, Arkansas Crop Improvement Program, Arkansas Agricultural Experiment Station, Fayetteville, AR 72704.

E-mail: rrobbin@comp.uark.edu

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